

Optimizing Cell-Free Protein Expression in the One-Pot PURE System

Insights into reaction composition and
translation efficiency

Yan Zhang¹, Matas Deveikis², Yanping Qiu¹, Lovisa Björn¹, Zachary A. Martinez¹, Tsui-Fen Chou¹, Paul S. Freemont², Richard M. Murray¹

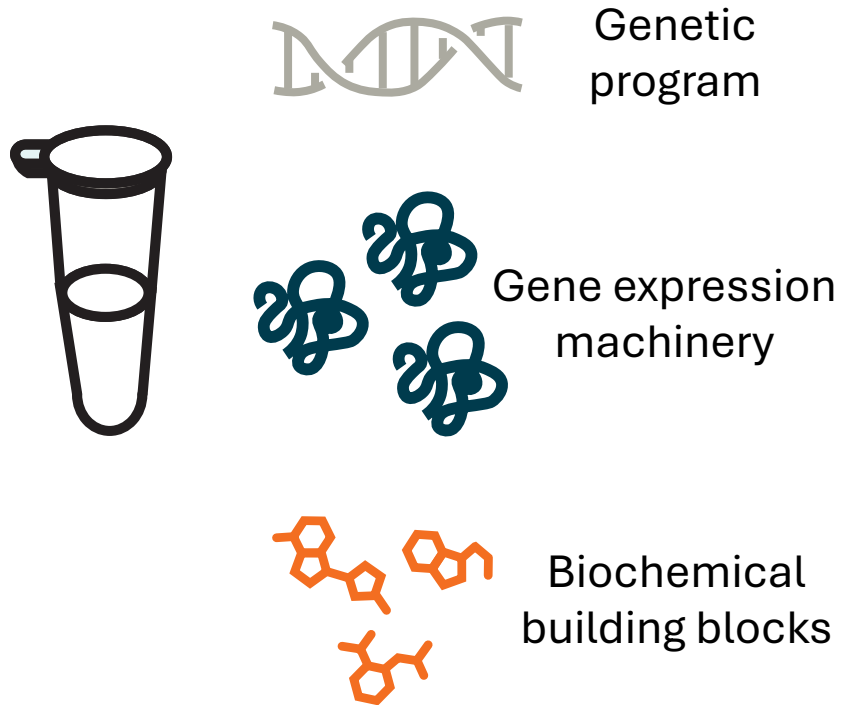
Presented at the 13th ICBE, January 7th, 2025, Houston, TX.

¹ California Institute of Technology | ² Imperial College London

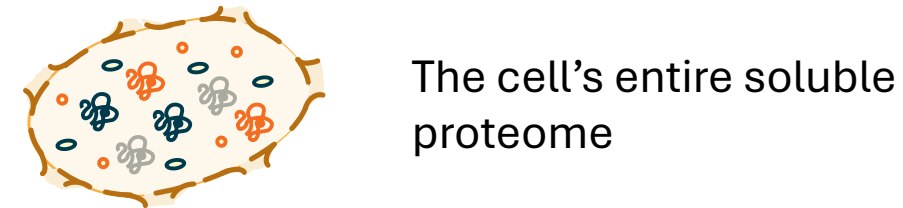
Caltech

Cell-free expression system is a powerful tool in breadboarding biomolecular designs

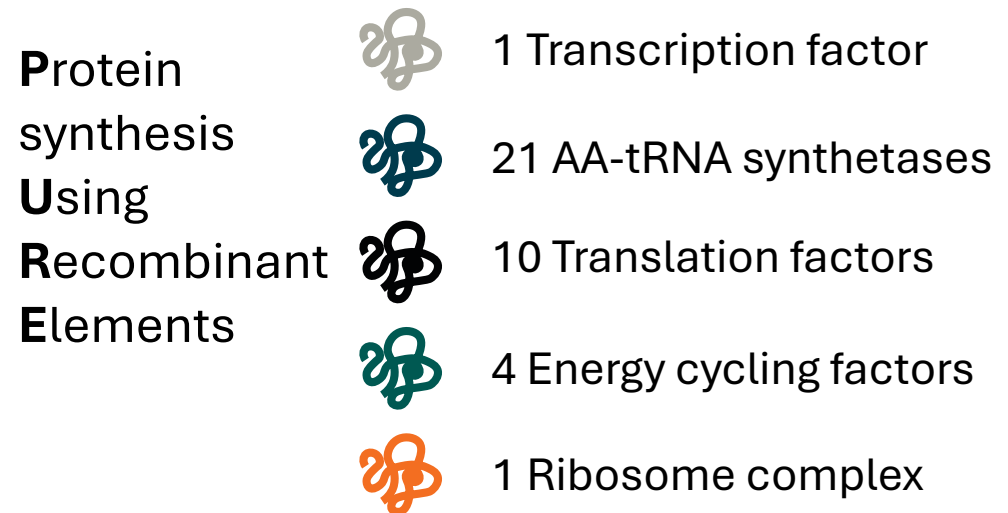
Cell-free expression system



Lysate-based cell-free system

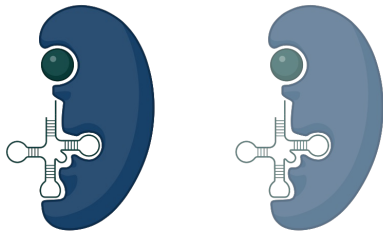


PURE-based cell-free system

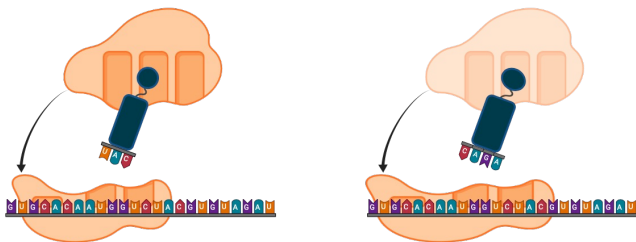


The PURE system can enable more biomolecular design possibilities over lysate-based cell-free systems

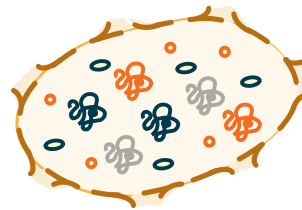
Replace aaRS for ncAA incorporation



Replace ribosome for quadruplet codon usage



Lysate-based cell-free system



The cell's entire soluble proteome

PURE-based cell-free system

Protein synthesis
Using Recombinant Elements



1 Transcription factor



21 AA-tRNA synthetases



10 Translation factors



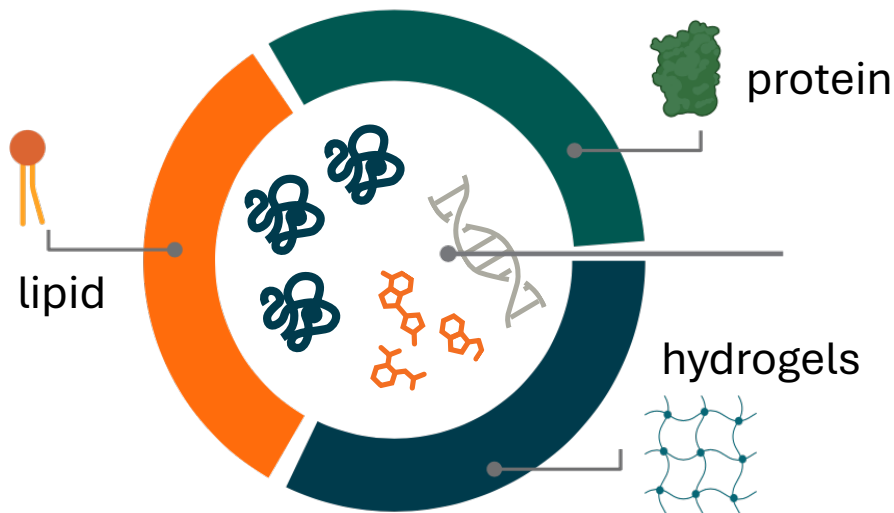
4 Energy cycling factors



1 Ribosome complex

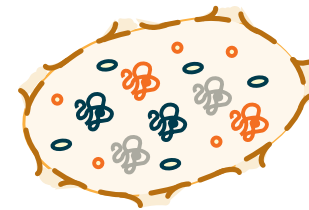
PURE system's minimal nature also makes it a desirable "cytosol" for synthetic cells

Synthetic cells – polymer container encapsulating biomolecules to execute cell-like functions



The minimal nature of PURE can be a desirable starting place to establish the "operating system" of synthetic cells






Lysate-based cell-free system



The cell's entire soluble proteome

PURE-based cell-free system






Protein synthesis Using Recombinant Elements

-  1 Transcription factor
-  21 AA-tRNA synthetases
-  10 Translation factors
-  4 Energy cycling factors
-  1 Ribosome complex

EXCEPT, making PURE system is highly labor-intensive

The One-Pot PURE system offers a streamlined solution

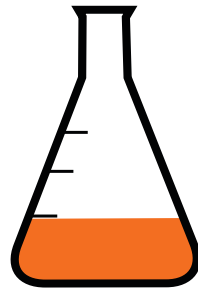
Machinery in the PURE system

-  1 Transcription factor
-  21 AA-tRNA synthetases
-  10 Translation factors
-  4 Energy cycling factors
-  Ribosome complex *
*purified separately

Traditional PURE

Shimizu *et al.*, 2001

Culture



Purify



Repeat for all
PURE proteins



~2 weeks to PURE

One-Pot PURE

Lavickova and Maerkl., 2019

Culture



Purify



In a single
preparation



~3 days to PURE

The One-Pot PURE appeared to be a better approach to set up synthetic cells for Yan's postdoc training plan



Imageflip.com, 2025

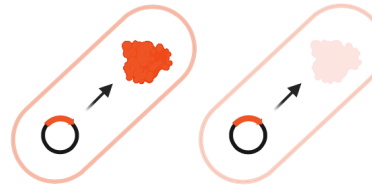
- Step 1:** Set up the One-Pot PURE system in the Murray Lab
- Step 2:** Encapsulate One-Pot PURE into synthetic cells
- Step 3:** Develop task-specialized synthetic cells
- Step 4:** Get results and publish

Overview of this talk:

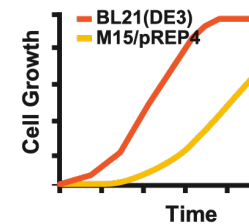
Unforeseen shortcomings with the One-Pot PURE system and mitigating strategies

The reaction's biochemical composition strongly influences the system's gene expression capacity

Loss of PURE protein expression



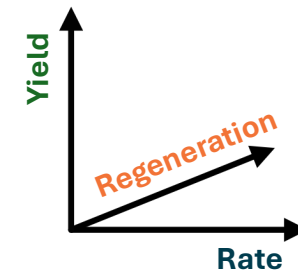
Slow growth and proteolysis in original expression strain



Some energy mixes are robust to a suboptimal PURE



A balanced reaction rate is key to higher protein yield

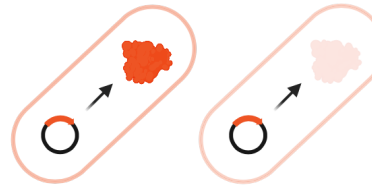


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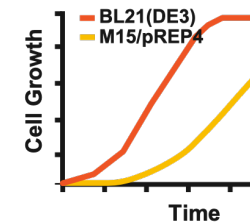
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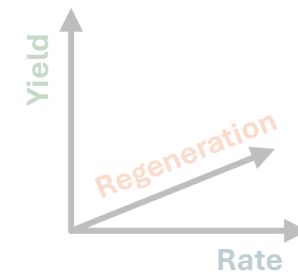
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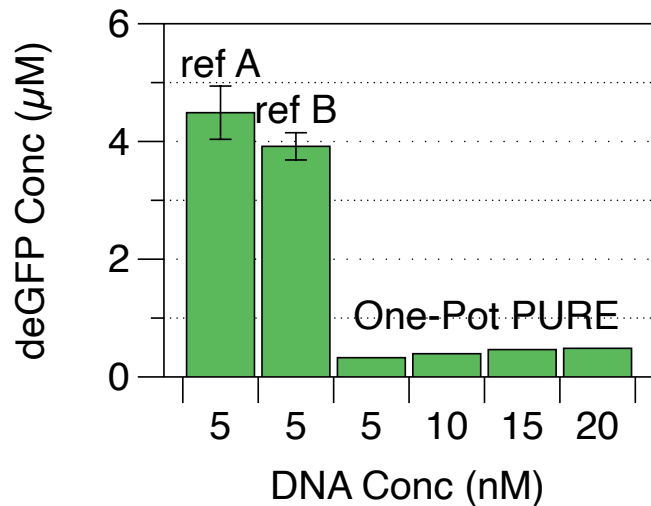
A balanced reaction rate is key to higher protein yield



Unstable PURE protein expression led to spontaneous protein dropouts in the co-culture

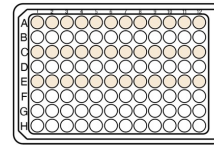


First batch of One-Pot PURE



The productivity of **One-Pot PURE** is **10 times lower** than that of commercial systems.

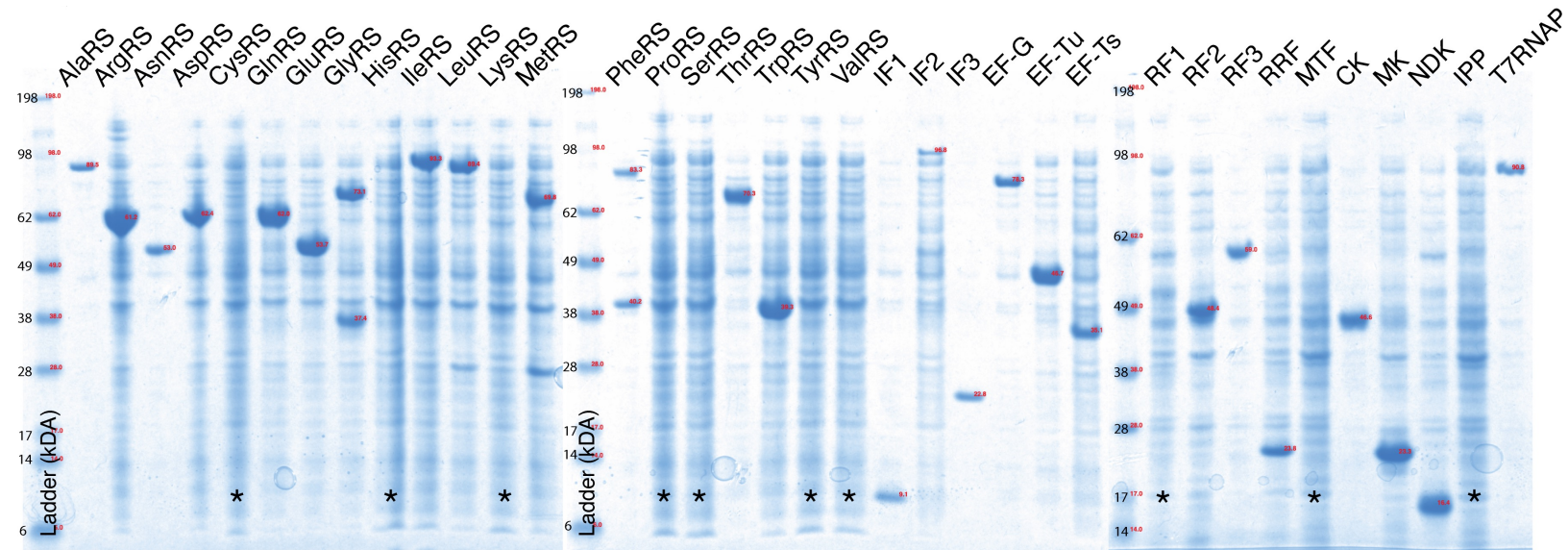
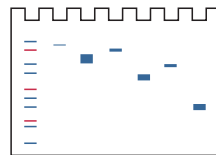
Grow cells in deep well plates



Induce protein expression w/ IPTG



Heat-denature cells to assess protein expression



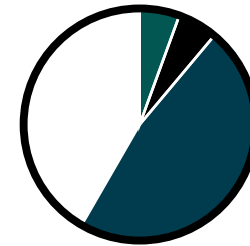
SDS-PAGE gel revealed that **multiple PURE proteins** lost visible expression. This drop-out may be the cause of low productivity.

Unstable PURE protein expression is caused by background expression causing growth burdens

Host distribution of PURE protein expression



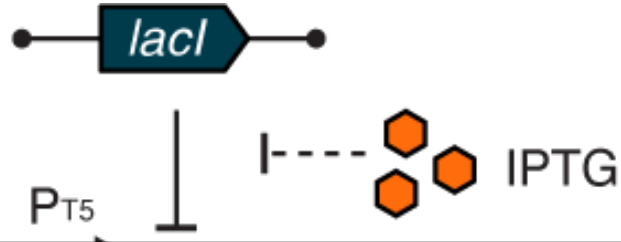
E. coli M15/pREP4
16/36 plasmids



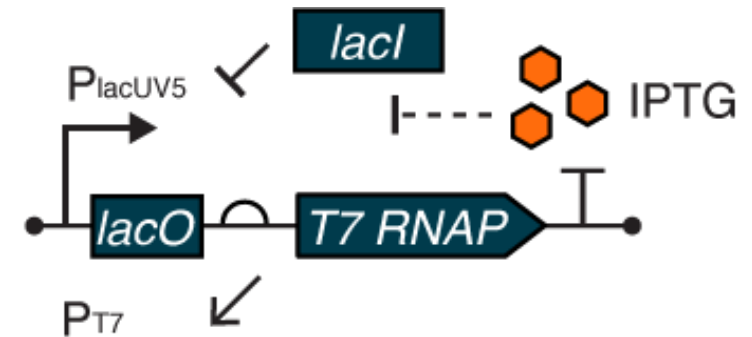
E. coli BL21(DE3)
20/36 plasmids

Expression regulation mechanism

pREP4



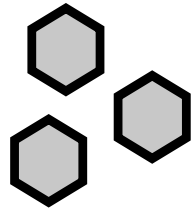
λDE3



pQE

In both cases, insufficient LacI would lead to burdensome background expression of PURE proteins, contributing to expression instability

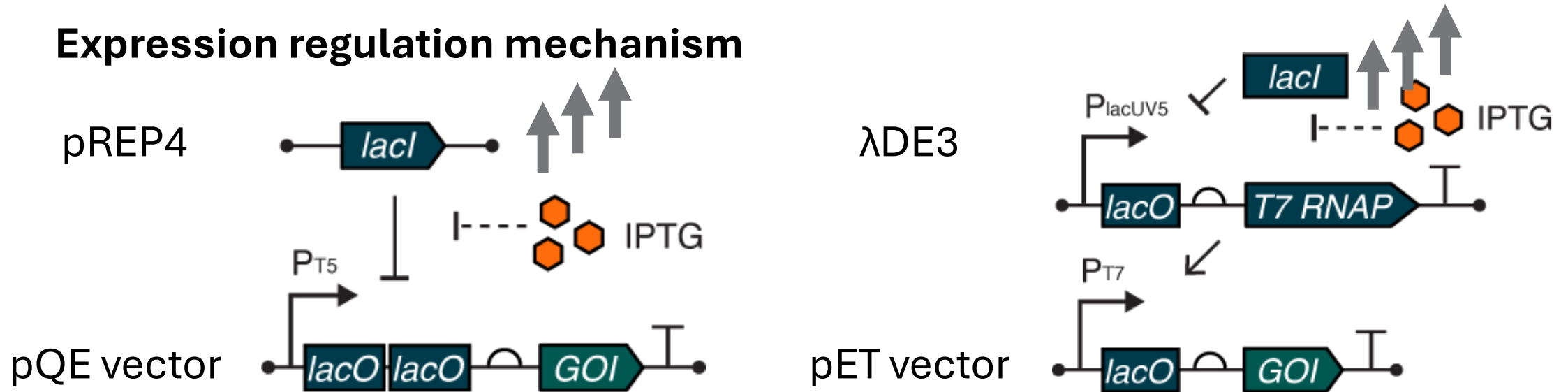
Adding glucose to overnight culture media improves expression stability via catabolite repression



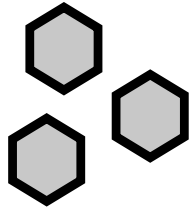
Glucose-mediated catabolite repression

In the presence of glucose, cells will upregulate repressors to turn off other carbon utilization pathways.

Expression regulation mechanism

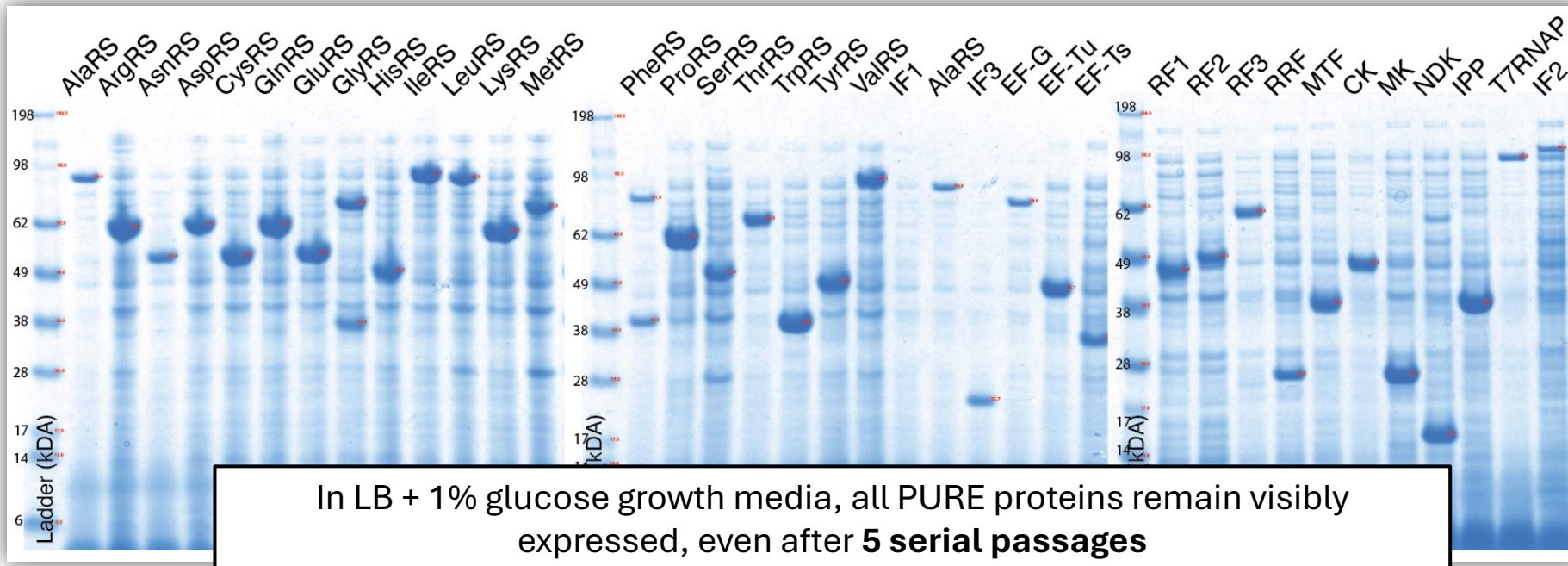


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Glucose-mediated catabolite repression

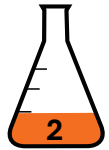
In the presence of glucose, cells will upregulate repressors to turn off other carbon utilization pathways.



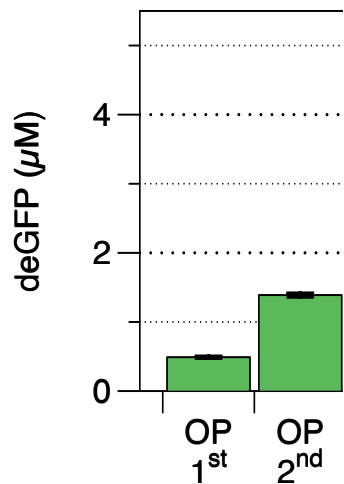
This is a common problem encountered by other labs.

Ngo *et al.* (2023) showed that expression stability can also be improved using BL21 Marionette strains carrying extra copies of *Lacl*.

Suboptimal PURE protein stoichiometry also leads to an unproductive PURE system

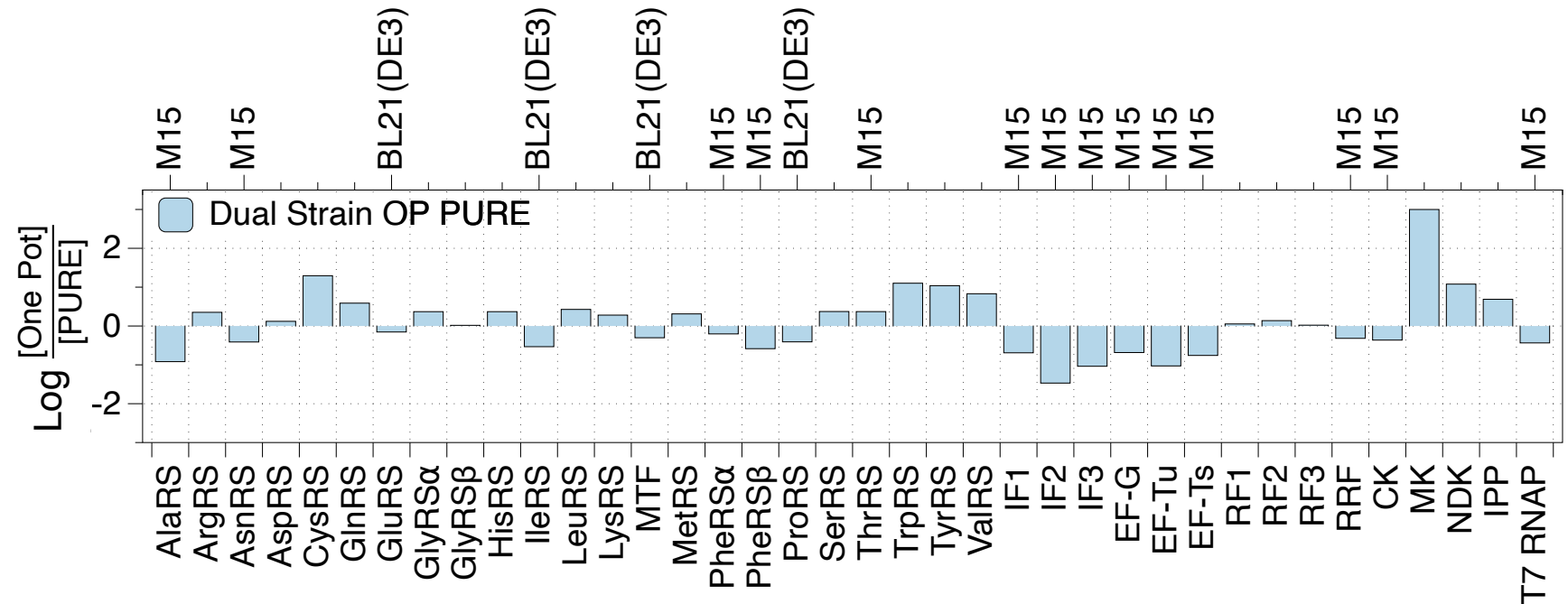


2nd attempt at
One-Pot PURE



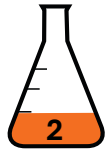
New batch is **3x more productive** than the previous batch.

But still **1/3 of the productivity** of commercial PURE

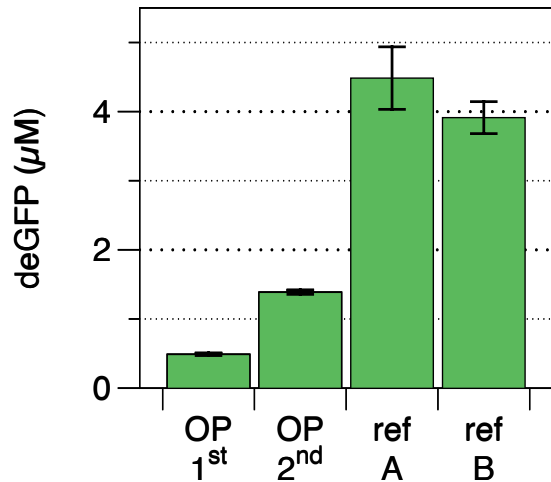


Mass spectrometry revealed the **abundance of many proteins in the One-Pot PURE mixture is lower** than that of commercial systems.

PURE protein deficiency may be correlated with slow *E. coli* M15 cell growth

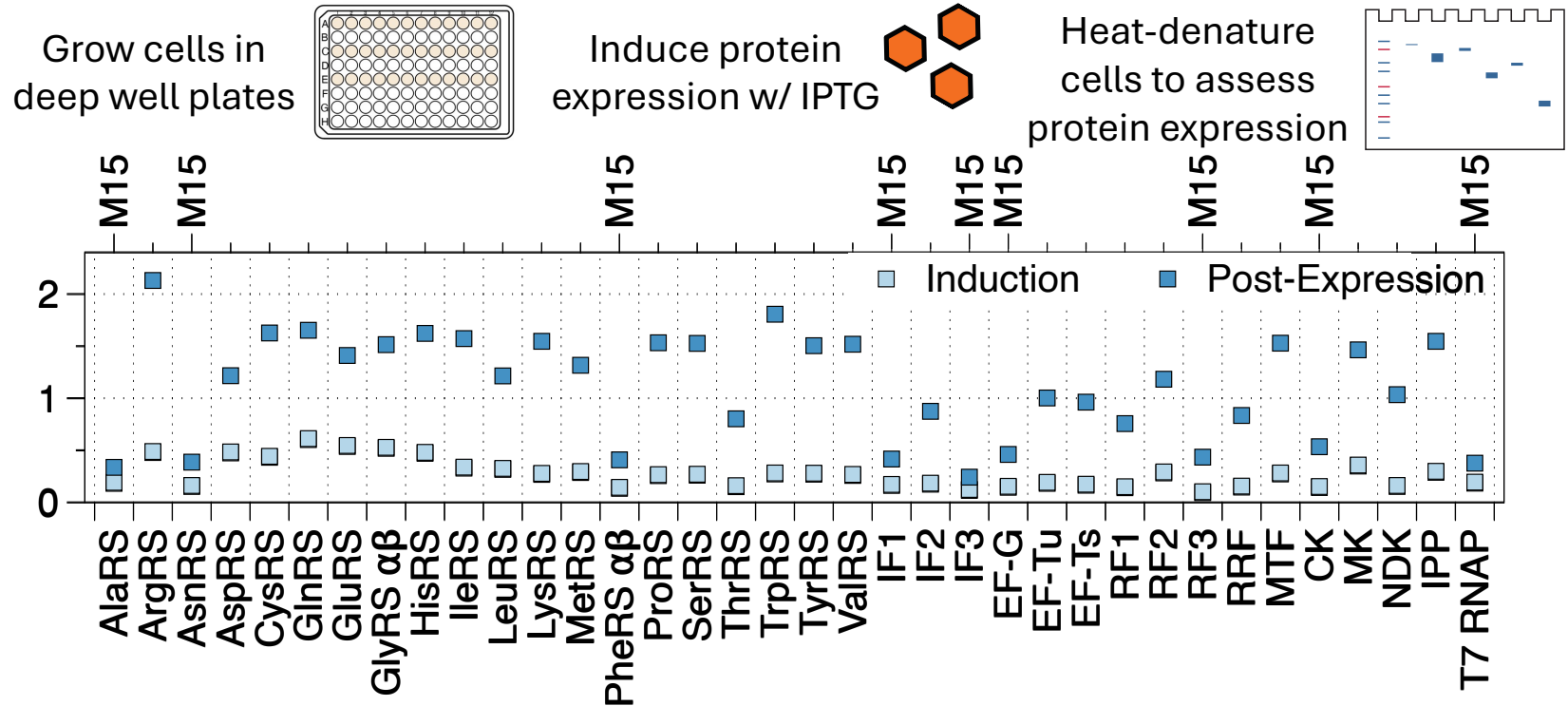


2nd attempt at
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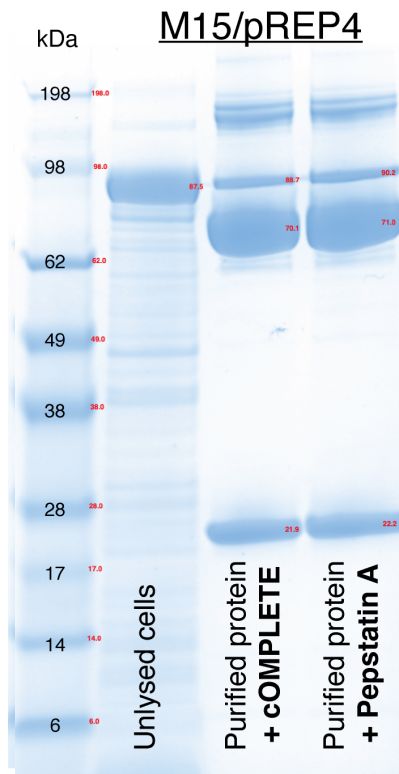
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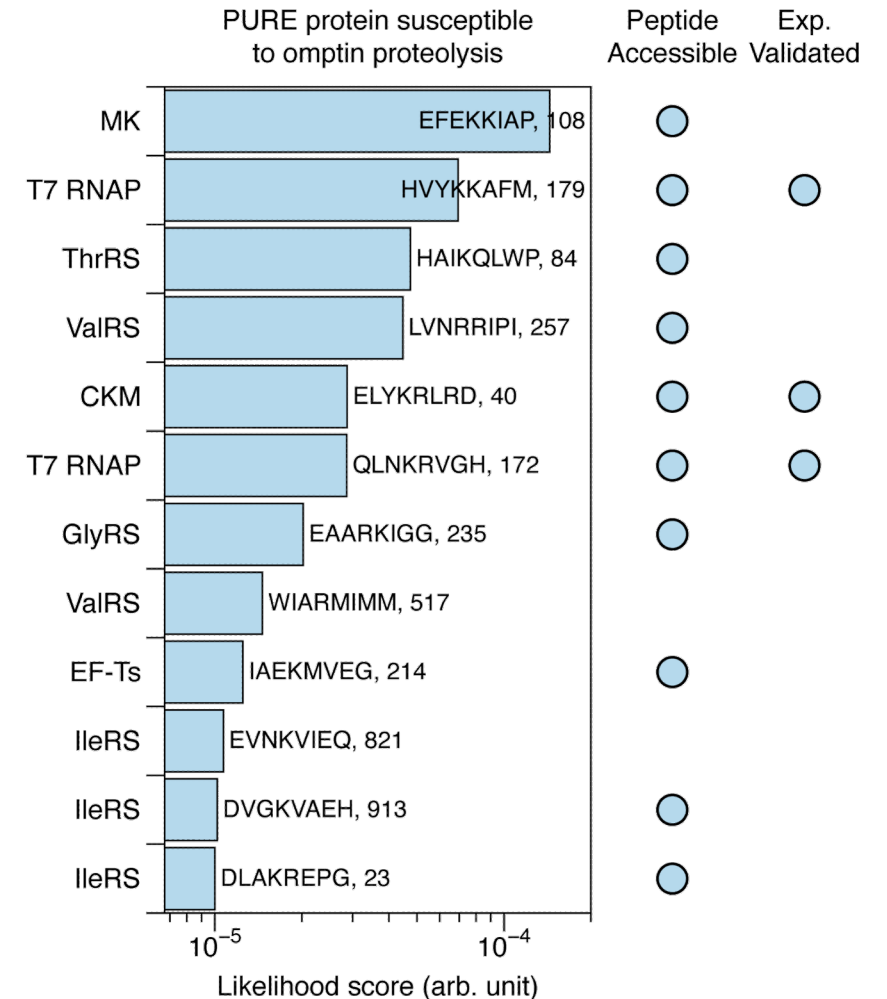
PURE proteins expressed in *E. coli* M15 consistently grow more slowly. The protein composition can be further reduced when grown in a co-culture.

E. coli M15 also exhibits omptin family proteolytic activities against PURE proteins

In purifying T7 RNAP from M15/pREP4, **proteolytic activity is observed at the purification step**, and this lysis is resistant to protease inhibitors.

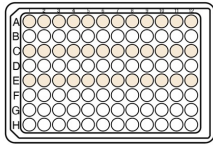


Based on OmpT's cleavage motifs, **at least 9 PURE proteins could be susceptible to OmpT proteolysis**



Expressing all PURE protein in BL21(DE3) led to more optimal protein stoichiometry and higher productivity

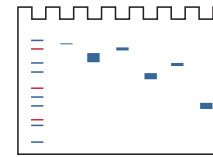
Grow cells in deep well plates



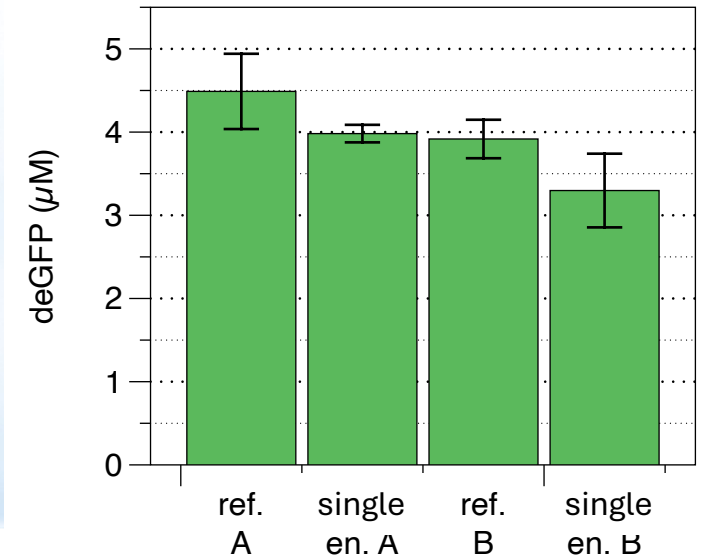
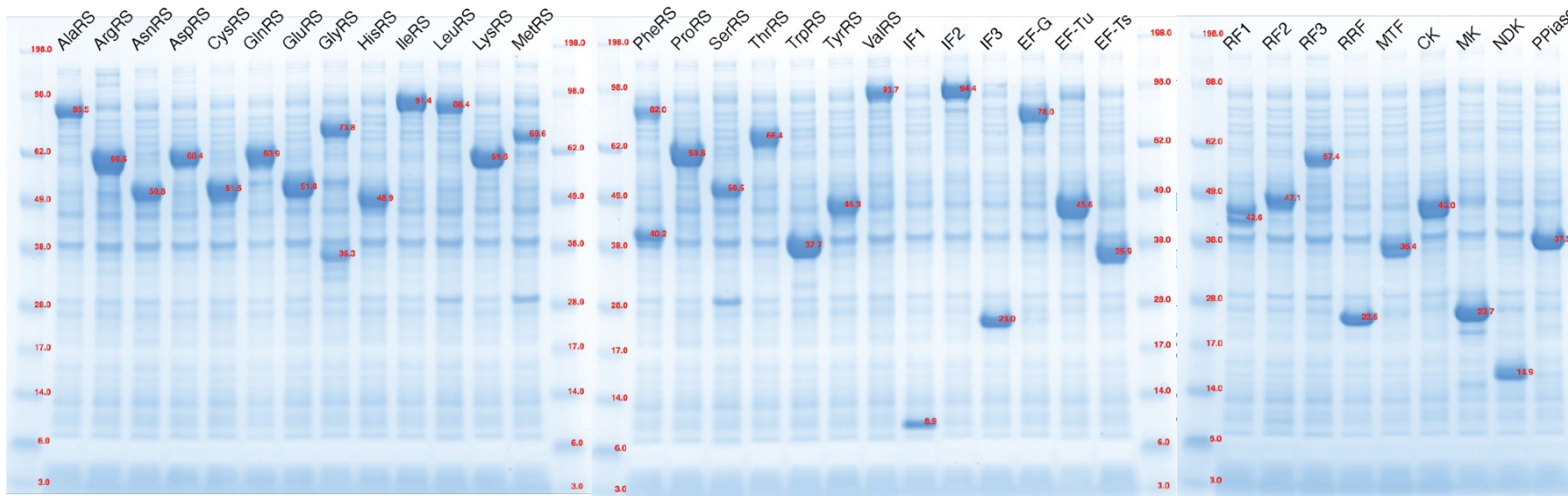
Induce protein expression w/ IPTG



Assess protein expression



Last attempt at One-Pot PURE



*we note the original PURE system (Shimizu *et al.*, 2001) expressed all PURE proteins in BL21/pREP4.

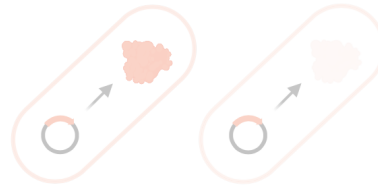
The BL21(DE3) ver. of One-Pot PURE is **within 15% of the protein productivity** compared to commercial systems.

Overview of this talk:

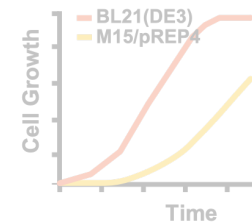
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Loss of PURE protein expression



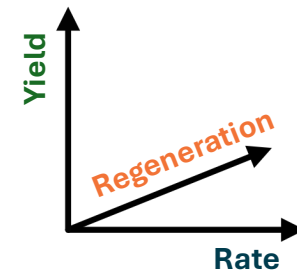
Slow growth and proteolysis in original expression strain



Some energy mixes are robust to a suboptimal PURE



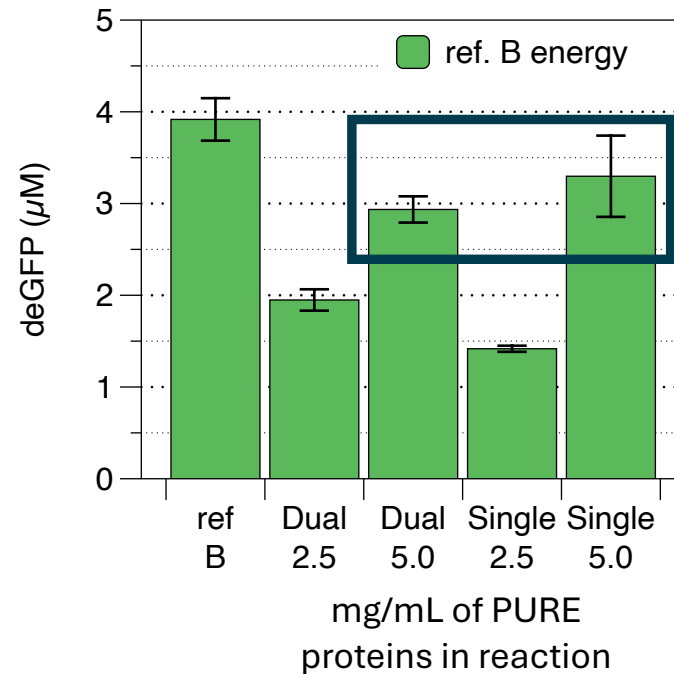
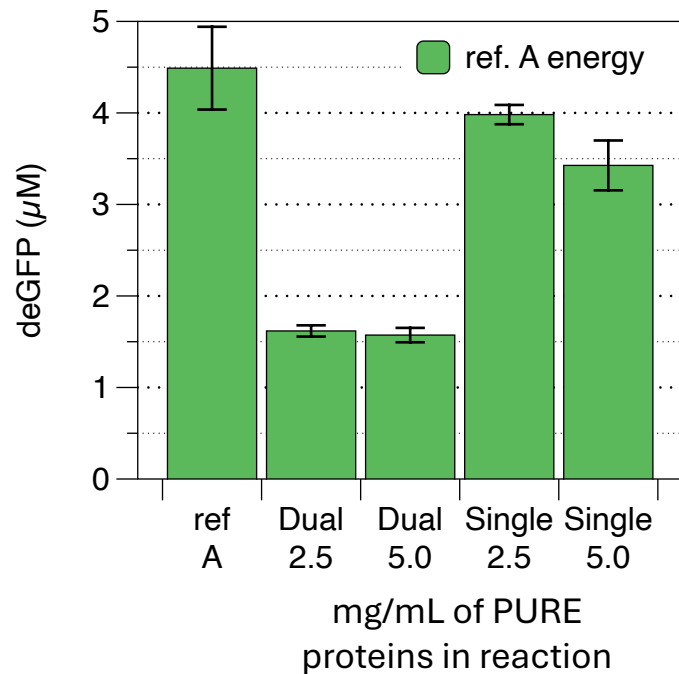
A balanced reaction rate is key to higher protein yield



Biochemical composition can play a compensatory effect on sub-optimal PURE protein stoichiometry

Different One-Pot PURE system behaviors observed in two commercial energy mixes

Dual = One-Pot PURE made by M15 and BL21(DE3)
Single = One-Pot PURE made by BL21(DE3)



Vendor B's **energy mix is robust to a sub-optimal PURE stoichiometry**

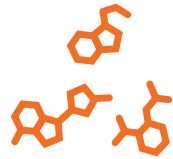
Biochemical building blocks (NTP, AA, etc.) of energy solutions alone **cannot explain the drastic difference**



Transfer RNAs (tRNA) may be a source of under-appreciated complexity in the energy solution

tRNA composition alone strongly influences the protein expression rate and the final reaction yield

Prepared the PURE energy mix in-house



Original Formulation

Varied the source of *E. coli* tRNA



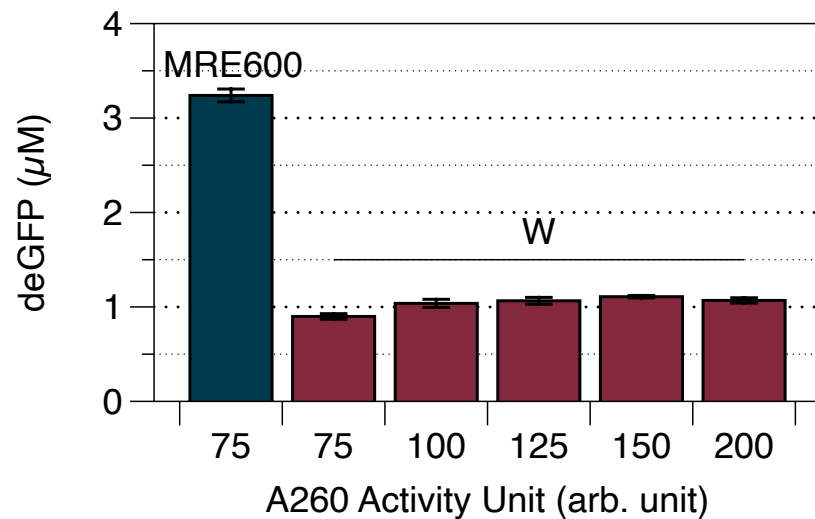
MRE600 Strain W

Added in the PURE protein mixtures



Single Strain OP

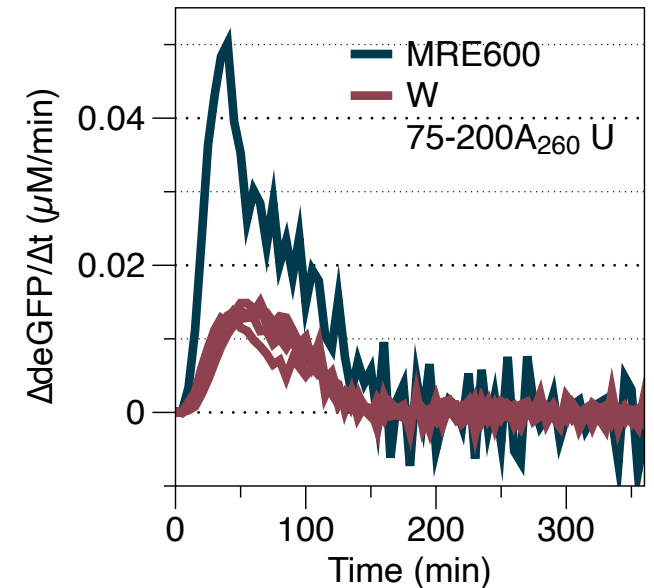
Comparing reaction yield



Reactions with **Strain W tRNA** are **3-fold less productive** compared to MRE600 tRNA.

*We note that the *E. coli* MRE600 tRNA from Roche was discontinued in 2022.

Comparing translation rate

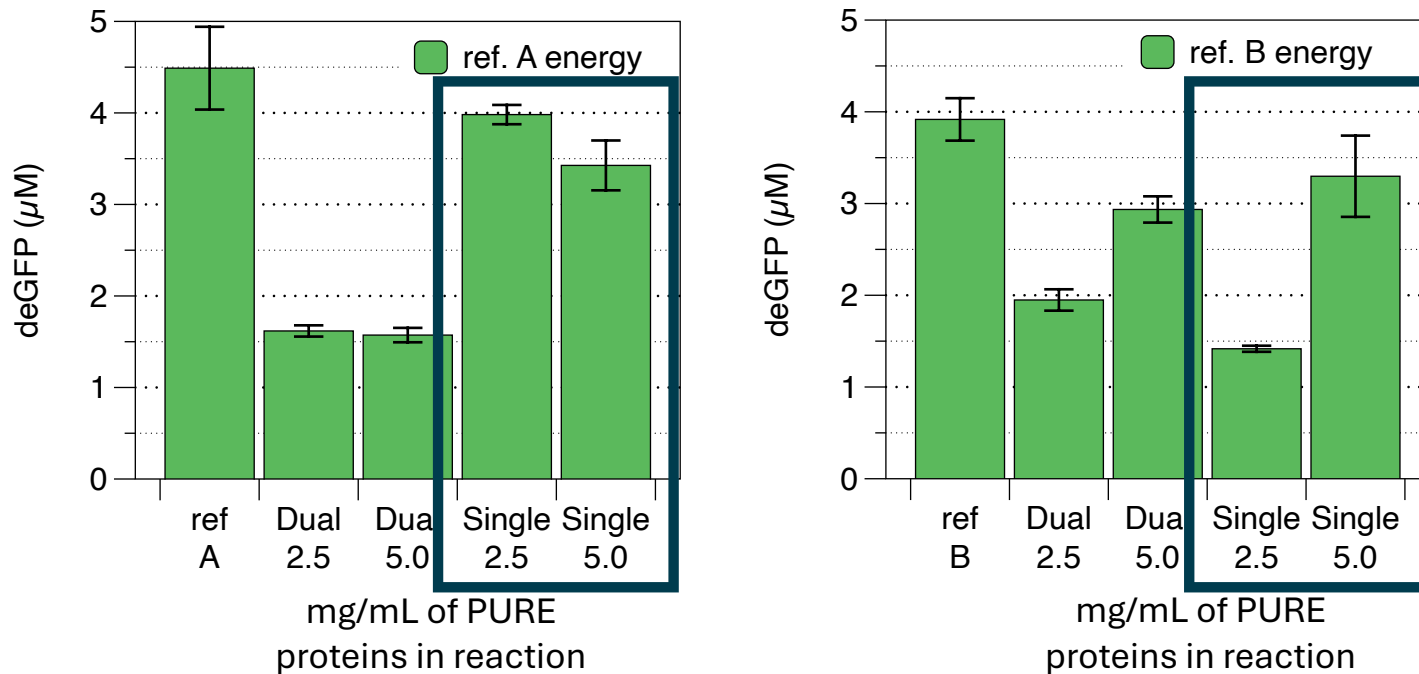


Strain W tRNA mix may have **deficient tRNAs, bottlenecking the reaction's translation capacity.**

High-yield protein expression requires a balanced protein production rate and energy formulation

Different One-Pot PURE system behaviors observed in two commercial energy mixes

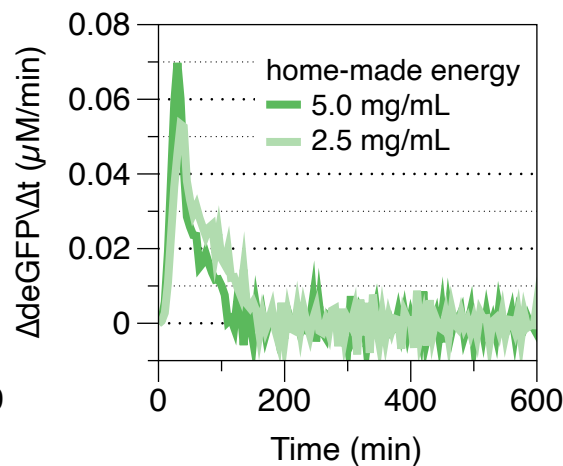
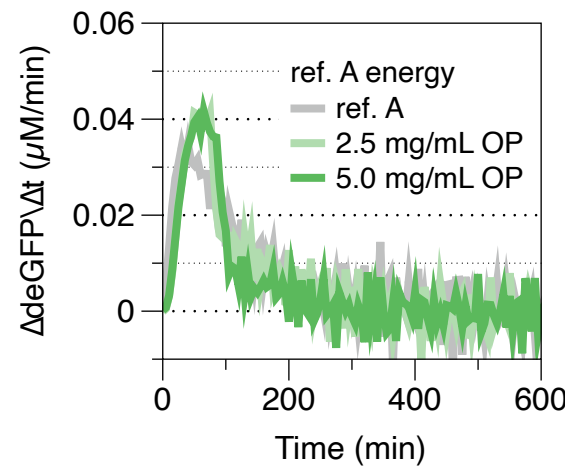
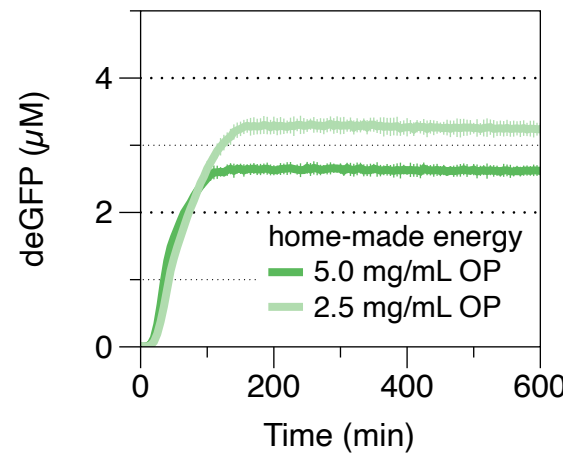
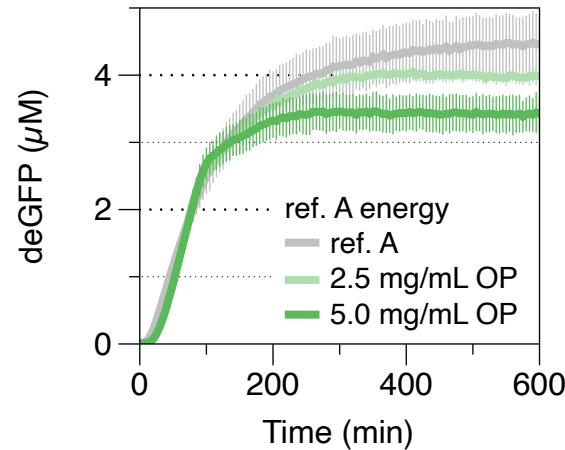
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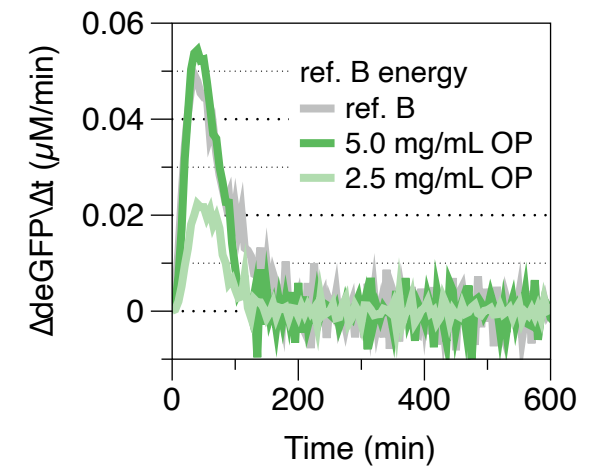
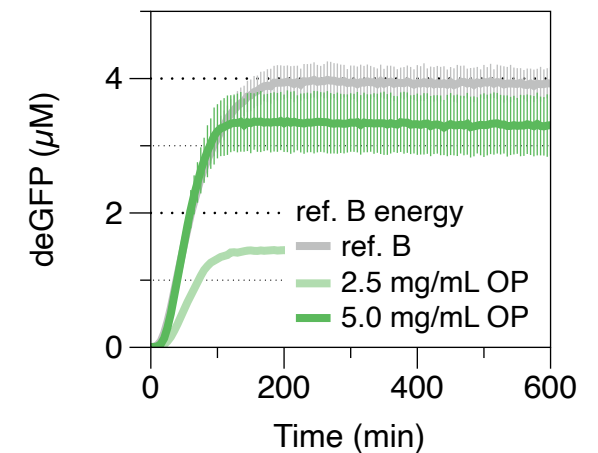
High-yield protein expression requires a balanced protein production rate and energy formulation

- Higher rate_{\max} is correlated with higher rate-drop
- Protein expression is energy-intensive. When the expression rate surpasses the system's energy regeneration rate, **ribosome stalling and premature termination** can happen
- But, the rate cannot be too low, where energy is spent without being productive

Too fast may be bad



Too slow can be worse

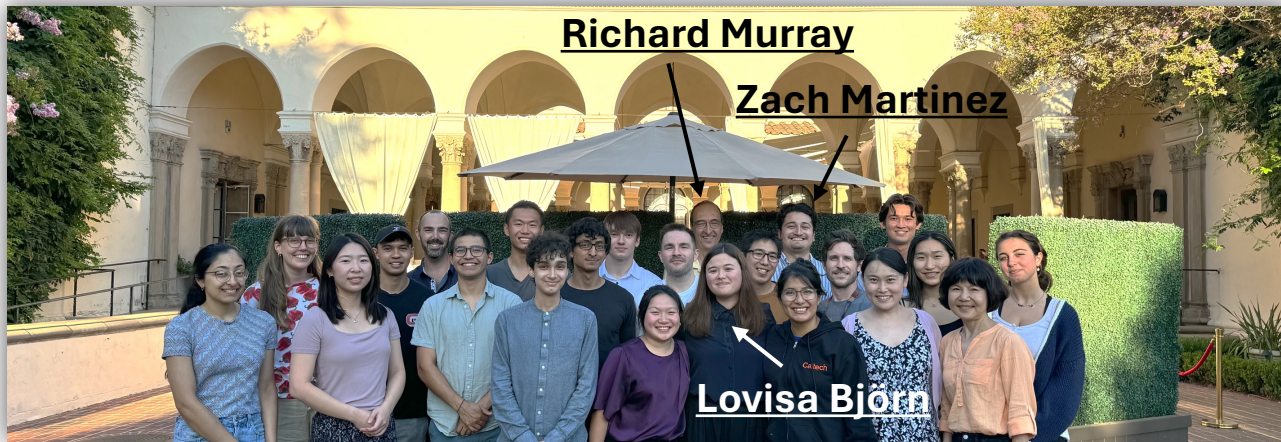


To set up a robust, productive One-Pot PURE

1. **Glucose-mediate catabolite repression** can be an effective strategy to improve genetic stability and prevent protein dropouts.
2. **E. coli M15 strain exhibits slower growth and proteolytic activity**, which is undesirable when making One-Pot PURE.
3. **The reaction's tRNA composition** strongly influences the protein expression capacity of PURE systems.
4. **High protein expression yield** in PURE systems requires balancing the expression and energy recycling rate.

Acknowledgment

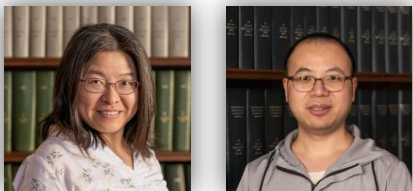
Murray Lab Members



Collaborators



Prof. Paul Freemont
and **Matas Deveikis**
at Imperial College
London



Prof. Tsui-Fen Chou
and **Dr. Eric Qiu**, at
Caltech Proteome
Exploration Laboratory

Funding Sources



Resources

Slides for
this talk

Manuscript
preprint



Plasmids developed in this work are
ready to distribute on Addgene